

# Myriapoda (Diplopoda, Chilopoda) of the Southern Cultures Park (Krasnodar Province, Southern Russia): unappreciated biodiversity

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## Abstract

The biodiversity of botanical gardens and arboreta is typically assessed by vegetation, birds, and insects such as beetles and butterflies. At the same time, most of the invertebrate animals live in litter and soil, while remaining hidden and poorly studied. Here we present the results of studies of two classes of myriapods in Southern Cultures Park, Adler, Krasnodar Province, Russia. During 2021, the diversity and abundance of myriapods were assessed by quantitative data obtained using pitfall traps. From these traps, 20 species of diplopods and 14 species of chilopods were recorded. Alpha diversity of myriapods was significantly higher ( $p=0.043$ ) in the spring-summer season in comparison to summer-fall. Beta diversity also significantly differed ( $p\leq 0.031$ ) between these seasons. The abundance of *Brachyiulus lusitanus* ( $p=0.018$ ), *Polydesmus mediterraneus* ( $p=0.047$ ), *Julus colchicus* ( $p=0.020$ ), and *Cylindroiulus placidus* ( $p=0.047$ ) was significantly higher in the spring season, while in the summer season, the abundance of Anthroleucosomatidae Gen. sp.1 ( $p=0.047$ ) was significantly dominant. Also, from 2018 to 2024, using hand collection from litter, 2 additional species of diplopods and 7 species of chilo-

Pods were recorded. Species of myriapods that were not previously recorded in Russia and introduced species were identified and illustrated.

### **Keywords**

Arboretum, centipedes, iconography, millipedes, new records, statistical ecology

## **Introduction**

It can be argued that botanical gardens and arboreta play a significant role in the sustainable development and conservation of plant biodiversity (Blackmore 2017, Ren and Blackmore 2023, Neves 2024). Furthermore, botanical gardens, which are home to a vast array of plants, play a pivotal role in the conservation of insects and spiders, both endemic to the region and invasive (Arteaga et al. 2020, Maynard et al. 2023). However, data on soil-dwelling arthropods remain deficient.

The Southern Cultures Park ("Yuzhnye Culture") is one of the oldest arboreta on the Black Sea coast and Russian part of the Caucasus. The park is a part of the Caucasian State Nature Reserve and is situated at the mouth of the Mzymta River, in the vicinity of Adler (43.417493°N; 39.935222°E), encompassing an area of 19.9 hectares. The plant collection is represented by 665 species, varieties, and garden forms (Soltani 2014, Shirayeva 2019). To date, studies of invertebrates inhabiting the territory of the arboretum have been limited to data on spiders. A total of 63 species of spiders have been recorded, with two species identified as invasive (Ponomarev et al. 2022).

Myriapods are soil invertebrates, comprising four classes. This study focuses on the two most prominent and species-rich classes: the centipedes, or Chilopoda, and the millipedes, or Diplopoda. Both groups of myriapods exhibit a high percentage of endemism in mountain ranges, including the Caucasus. Chilopods are predatory and may consume insects, nematodes, annelids, molluscs, and others, thereby influencing the population dynamics of their prey. In contrast, diplopods primarily consume dead plant matter, occasionally shifting to feeding on living plants and becoming pests (Minelli 2011, 2014, Minelli and Golovatch 2024).

The present paper presents the results of studies of chilopods and diplopods in Southern Cultures Park, including their species composition and abundance dynamics.

## **Materials and methods**

### **Sampling methods**

The collection of material was carried out using pitfall traps (eg Siewers et al. 2014) in 2021 and hand sampling under bark, stones, and litter in 2018 and 2021–2023. At the study site were placed 15 traps (500 ml plastic glasses with 4% formaldehyde

solution). Traps were installed on March 25 and checked on April 06, April 28, May 25, June 17, July 14, July 29, August 20, and September 18. Selected myriapods were fixed in 70% alcohol for further identification.

## Imaging

The habitus photographs were taken with a Zeiss StereoDiscovery V.20 microscope using focus-stacking technology; the final image was compiled from multiple layers using ZEN Software. The photographs of chilopods were taken with a Levenhuk D800T digital camera; the final image was compiled from multiple layers using Helicon Focus 8.2.2 Software.

Scanning electron micrographs of the gonopods and other taxonomically significant characters were taken with a Zeiss CrossBeam 340 scanning electron microscope (SEM) (Rostov-on-Don State Technical University, Rostov-on-Don, Russia). For some SEM micrographs, the objects were placed on an SEM-stub and air dried for two days in a glass filled with Silica gel. After examination, material was removed from stubs and returned to alcohol.

All images were processed using Adobe Photoshop 2020 software.

## Statistical analysis

The programming language R v4.4.0 (R Foundation for Statistical Computing, Vienna, Austria) was used for statistical analyses. The “vegan” package (version 2.6-6.1) was used for the calculation of alpha and beta diversity indices (Dixon 2003). For the comparison of the alpha diversity of myriapods in Southern Cultures Park, we used the Shannon index (Shannon and Weaver 1964), the Simpson index (Simpson 1949), and the Pielou index (Pielou 1966). For the beta diversity analysis, we used the Bray–Curtis dissimilarity (Sorenson 1948) and Jaccard similarity (Jaccard 1908) indices. For statistical comparison, observation periods were grouped as follows: March 25–June 16 (spring season), and June 17–September 18 (summer season). For the comparison of the alpha diversity indices and the abundance of studied myriapods, the Mann–Whitney test was used. PERMANOVA with the “adonis” function from the “vegan” package was used to determine differences in beta diversity distances (the number of permutations was set to 999). P-values were considered significant at  $p < 0.05$ . Results were visualized with the “ggplot2” package (Wickham 2016).

## Results

### Species diversity

As a result of sampling using pitfall traps in the Southern Cultures Park, 20 species of diplopods and 14 species of chilopods were recorded (Suppl. material 1: Table).

Also, 9 millipede and 13 centipede species were collected by hand sampling under bark, stones, and litter. Some species have only been defined to the genus or family level due to the fact that only females have been found in the case of diplopods, or undescribed species in the case of chilopods.

Of all the myriapod species found, 7 have a cosmopolitan or subcosmopolitan distribution:

*Brachyiulus lusitanus* Verhoeff, 1898 (Fig. 1) – an ubiquitous anthropochoric species (Kime and Enghoff 2017, Vagalinski and Lazányi 2018). It is an invasive species in the Caucasus, previously recorded in Abkhazia and Georgia, and is formally new to Russia (Vagalinski and Golovatch 2021).

*Cylindroiulus truncorum* (Silvestri, 1896) (Fig. 2) – a subcosmopolitan species, probably of North African origin. Found in botanical gardens, greenhouses, parks and other synanthropic habitats (Kime and Enghoff 2017, Stoev et al. 2010). Invasive in the Caucasus, previously found in a greenhouse in Rostov-on-Don, Ciscaucasia (Evsyukov and Golovatch 2013).

*Nopoiulus kochii* (Gervais 1847) – a cosmopolitan species, probably of Caucasian origin (Kime and Enghoff 2017, Golovatch and Enghoff 1990).

*Oxidus gracilis* (C.L. Koch, 1847) (Fig. 3) – a species of East Asian origin, widely distributed in greenhouses and botanical gardens all over the world, where it probably spread with planting material (Stoev et al. 2010, Kime and Enghoff 2011, Evsyukov et al. 2016). It has been successfully naturalised on the Black Sea coast of the Caucasus and is also found in natural habitats.

*Lamyctes africanus* (Porat, 1871) (Fig. 6) – a cosmopolitan species known from Africa, Australia, Europe and anthropogenic habitats in Siberia (Nefediev et al. 2020). The first record for the Caucasus. The genus *Lamyctes* belongs to the family Henicopidae, which is mainly distributed in the southern hemisphere, but some of its representatives are anthropochore species and are extremely widespread (Minelli 2011).

*Lamyctes coeculus* (Brölemann, 1889) (Fig. 7) – a cosmopolitan, found in Europe, Australia, North and South America, Africa, the Middle East and Siberia (Nefediev et al. 2016, Gilgado et al. 2022). This is the first record in the Caucasus.

*Lithobius forficatus* (Linnaeus, 1758) – an anthropochore species of European origin that has been introduced to the Middle East, the eastern Palearctic, Australia, North and South America, and Africa (Nefediev et al. 2016). In the Caucasus, it is restricted mainly to steppe and anthropogenic habitats (Zuev 2016).

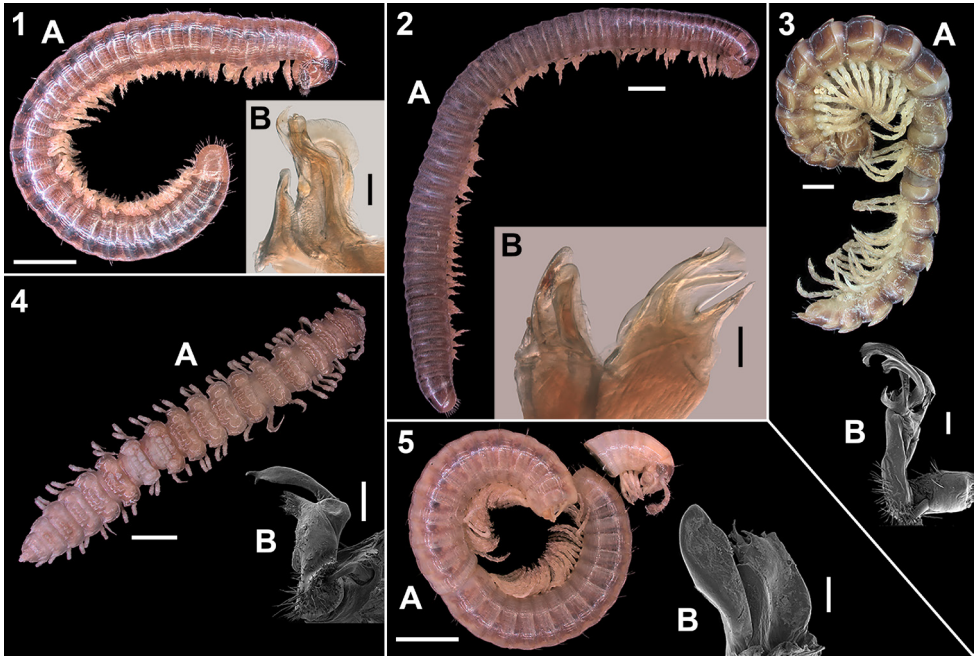
One species has a transpalearctic chorotype:

*Pachymerium ferrugineum* (C.L. Koch, 1835), a eurybiont species widely distributed in the Caucasus (Dyachkov et al. 2022).

Five species have Mediterranean chorotypes:

*Lophoproctus coecus* Pocock, 1894 – distributed throughout Europe to Central Asia, widely presented in the Caucasus, including Krasnodar Province (Short 2015).

*Polydesmus mediterraneus* Daday, 1889 (Fig. 4) – Eastern Mediterranean species, probably introduced to the Caucasus. New record for Russia, previously recorded in Abkhazia (Kime and Enghoff 2011, Golovatch et al. 2016).



**Figures 1–5.** Habitus (A) and gonopods (B) of some millipedes, lateral views: 1 – *Brachyiulus lusitanus* Verhoeff, 1898, 2 – *Cylindroiulus truncorum* (Silvestri, 1896), 3 – *Oxidus gracilis* (C.L. Koch, 1847), 4 – *Polydesmus mediterraneus* Daday, 1889, 5 – *Omobrachiulus hortensis* (Golovatch, 1981). Scale bars: 10 mm (A), 0.1 mm (B).

*Clinopodes escherichii* (Verhoeff, 1896) – Eastern Mediterranean species, known in the Caucasus from Stavropol Province, Dagestan and the Black Sea coast (Korobushkin et al. 2016, Dyachkov et al. 2022).

*Diphyonyx conjungens* (Verhoeff, 1898) – Eastern Mediterranean species, widely distributed in the Caucasus (Dyachkov et al. 2022).

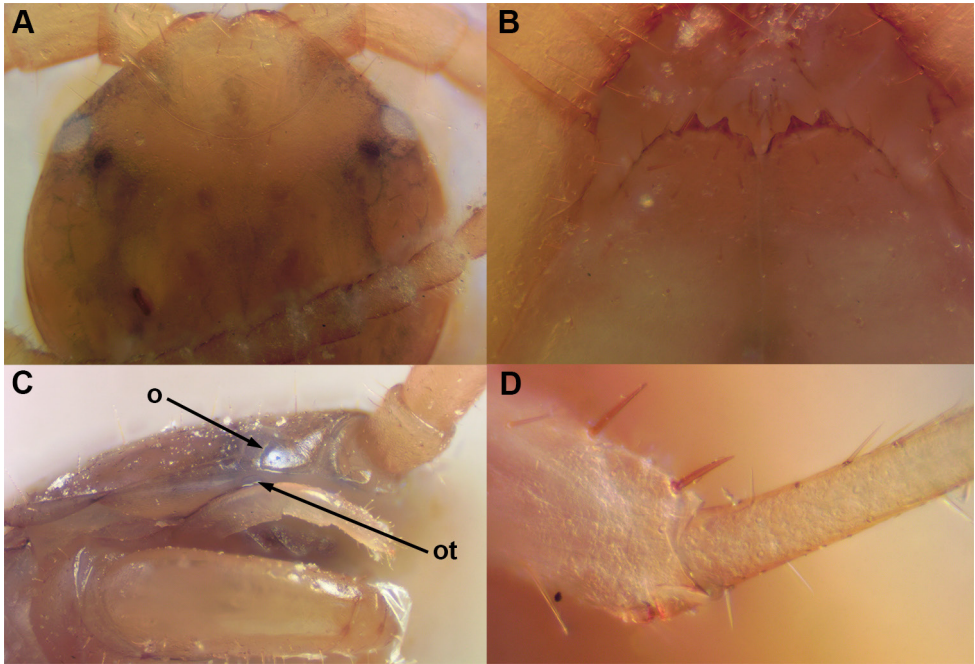
*Lithobius peregrinus* Latzel, 1880 – Eastern Mediterranean species, widespread in the Caucasus (Zuev 2016).

Seven species are mainly distributed in Europe:

*Clinopodes flavidus* C.L. Koch, 1847 (Fig. 8) – distributed mainly in central and eastern Europe, apparently an anthropochore species in the Caucasus (Bonato et al. 2011).

*Geophilus carpophagus* Leach, 1816 (Fig. 9) – European species, probably introduced. A new species for the Caucasus, from the territory of Russia there was previously a dubious finding in the Ulyanovsk Region (Volkova 2016).

*Geophilus* cf. *flavus* De Geer, 1778 – a European species introduced to North America and Siberia (Nefediev et al. 2017). In the Caucasus, it is known from the Krasnodar and Stavropol Provinces (Zuev 2016), probably also an introduced species.



**Figure 6.** *Lamyctes africanus* (Porat, 1871): head, dorsal view (A), anterior part of forcipular segment, ventral view (B), head, lateral view (C), tibia 12, distal spinose projection (D). Taken not to scale. Abbreviations: o – ocellus; ot – organ of Tömösvary.

*Henia illyrica* Meinert, 1870 – a southeastern European species, known in the Caucasus from Krasnodar and Stavropol Provinces (Zuev 2016).

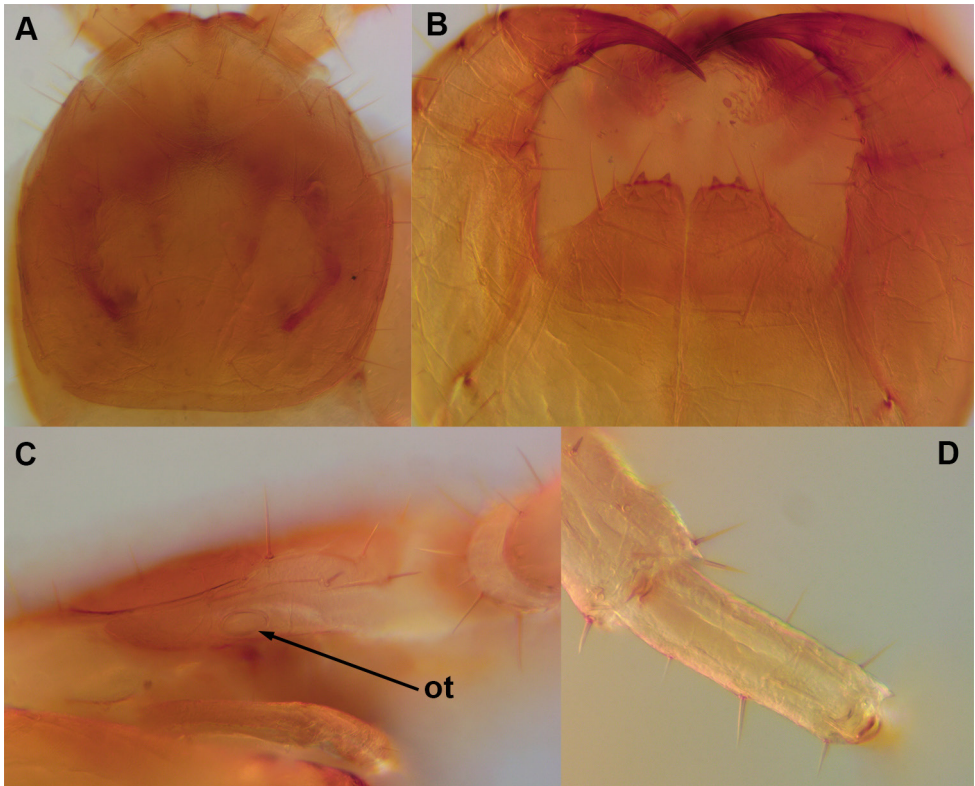
*Harpolithobius* cf. *anodus* Latzel, 1880 (Fig. 10) – a south-eastern European species early recorded only from Krasnaya Polyana, Krasnodar Province (Lignau 1903), probably an introduced species. The specimens examined differ from the description of this species by the presence of a small armed dorsodistal swelling on tibia 15 of the male.

*Lithobius mutabilis* L. Koch, 1862 – European species, distributed in the Caucasus (Korobushkin et al. 2016).

*Lithobius sselivanoffi* Garbowski, 1897 – probably south-eastern European species, the exact distribution is unknown due to confusion in the description. Widespread in the Caucasus (Zalesskaja 1978).

Another species, *Clinopodes* cf. *latisternus* (Attems, 1947) (Fig. 11), was previously known only from two specimens from southern Anatolia (Bonato et al. 2011), and its distribution and biology are virtually unknown. This is the first record of this species in the Caucasus and Russia.

The remaining species are endemic and subendemic to the Caucasus. Of these, only one species is recorded for the first time in Russia:



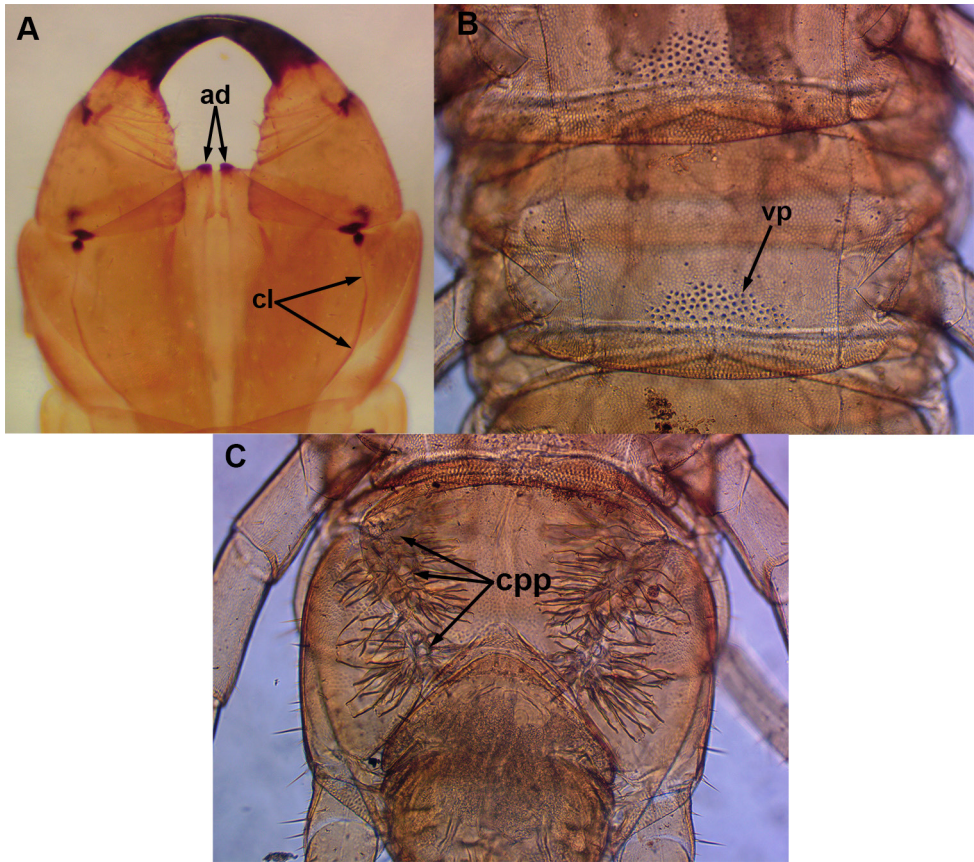
**Figure 7.** *Lamyctes coeculus* (Brölemann, 1889): head, dorsal view (A), anterior part of forcipular segment, ventral view (B), head, lateral view (C), tibia 12, distal spinose projection (D). Taken not to scale. Abbreviation: ot – organ of Tömösvary.

*Omobrachiulus hortensis* (Golovatch, 1981) (Fig. 5) – endemic to the Colchidan part of the Caucasus, previously recorded in Abkhazia (Vagalinski and Golovatch 2021). The most northeastern locality for the species and new record for Russia.

Two undescribed chilopod species are probably endemic to the Caucasus:

*Harpolithobius* sp. was represented by a single female. Based on the combination of the following characters: 2+2 denticles on the forcipular coxosternite, presence of spurs on the first pair of walking legs, flat and broad tarsungulum of the forcipules, simple claws of the female gonopods (Fig. 12), we cannot assign it to the known representatives of the genus and more material is needed to describe the new species.

*Lithobius* sp. is close to *L. curtipes* C.L. Koch, 1847, but there are some differences: a smaller dorsodistal projection on the 15th tibia of males and the presence of an additional claw on the 15th pair of legs (Fig. 13). Further studies are needed to clarify the taxonomic status of this species.



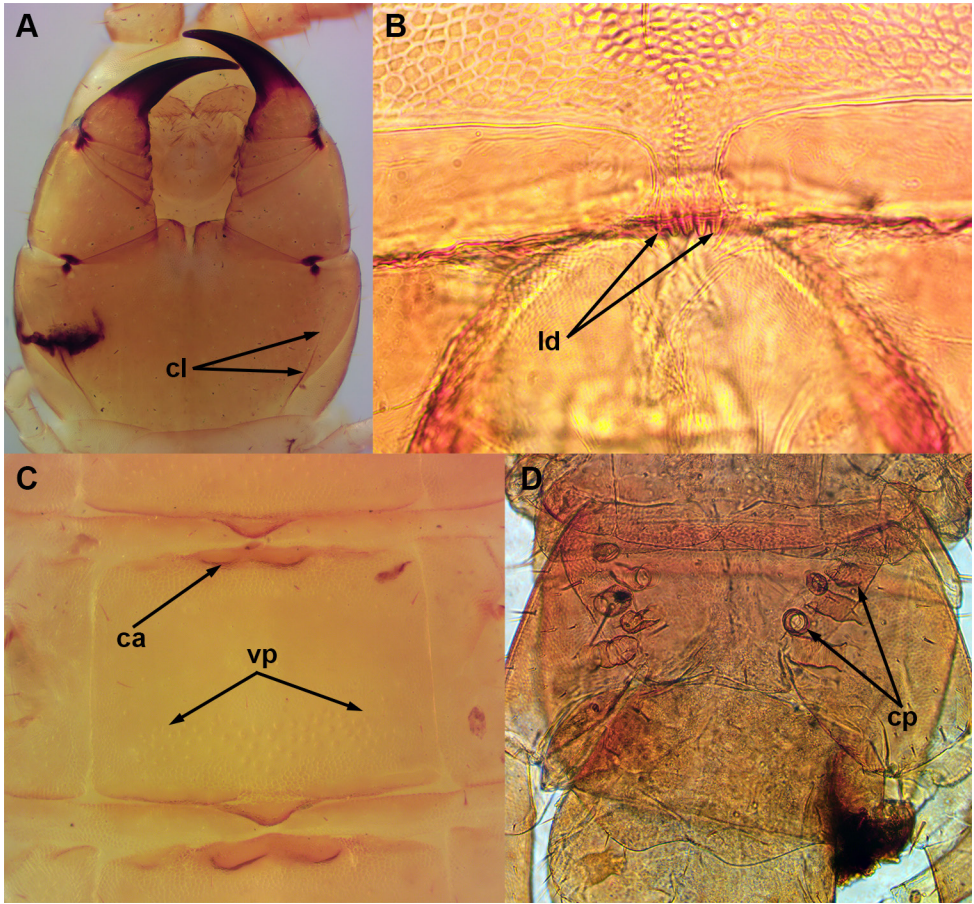
**Figure 8.** *Clinopodes flavidus* C.L. Koch, 1847: forcipular segment (A), metasternites of the most posterior leg-bearing segment (B), ultimate leg-bearing segment (C), ventral views. Taken not to scale. Abbreviations: ad – anterior denticles, cl – chitin-lines, cpp – coxopleural pit, vp – ventral pore field.

### Diversity and abundance of myriapods in 2021

There were 34 species of myriapods according to the data obtained using pitfall traps between March 25, 2021, and September 18, 2021 (Suppl. material 1: Table). Among dominant species, there were *Brachydesmus kalischewskyi*, *Harpolithobius* cf. *anodus*, *Brachyiulus lusitanus*, *Lophoproctus coecus*, *Lithobius striatus monosulcatus*, *Polydesmus mediterraneus*, *Trachysphaera radiosa*, *Clinopodes escherichii* and others (Fig. 14).

Alpha diversity values, Shannon and Simpson index, in particular, were significantly lower in the summer season ( $p=0.043$ ). However, there were no significant differences in Pielou indices, which indicates that the evenness of the Myriapoda species in the studied area is not affected by seasonal changes (Fig. 15).



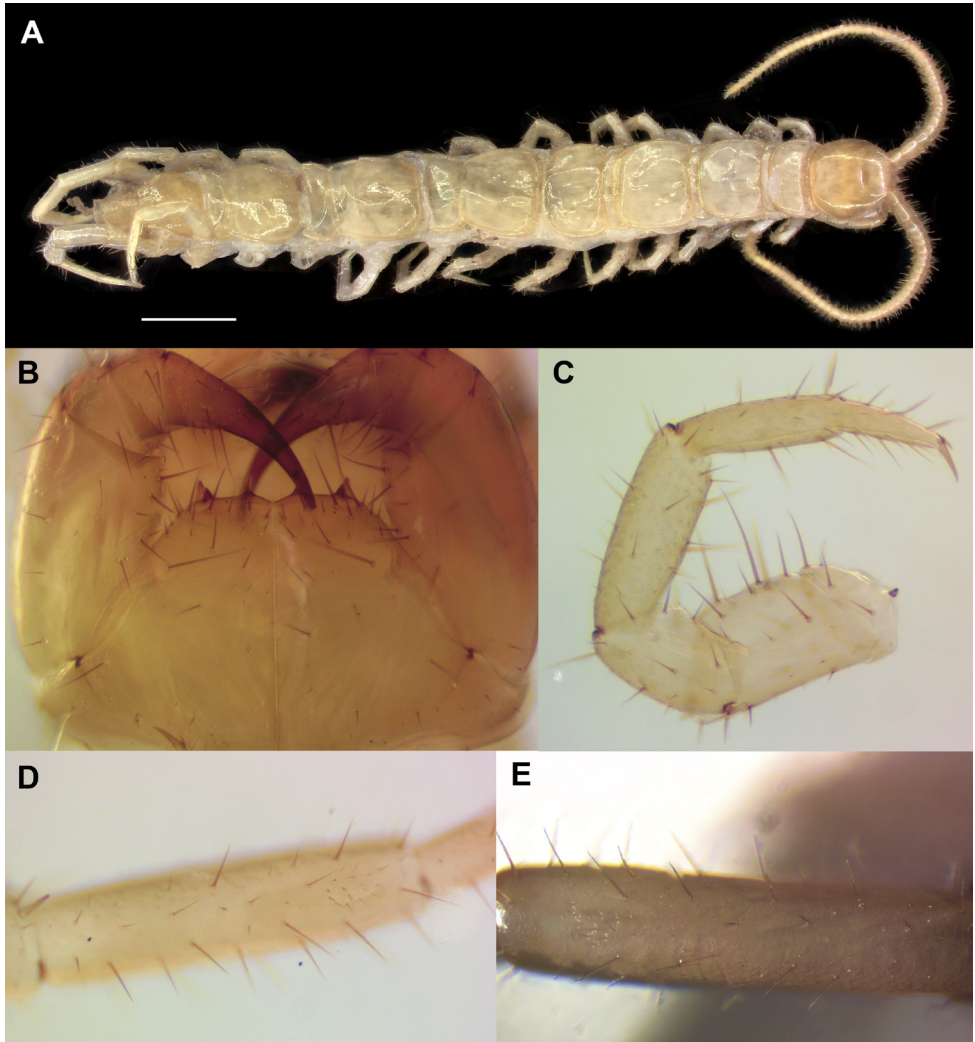


**Figure 9.** *Geophilus carpophagus* Leach, 1816: forcipular segment (A), labrum (B), metas-  
 ternite of 5th leg-bearing segment (C), ultimate leg-bearing segment (D), ventral views.  
 Taken not to scale. Abbreviations: ca – carpophagus pit, cl – chitin-lines, cp – coxal pores,  
 ld – labral denticles, vp – ventral pore field.

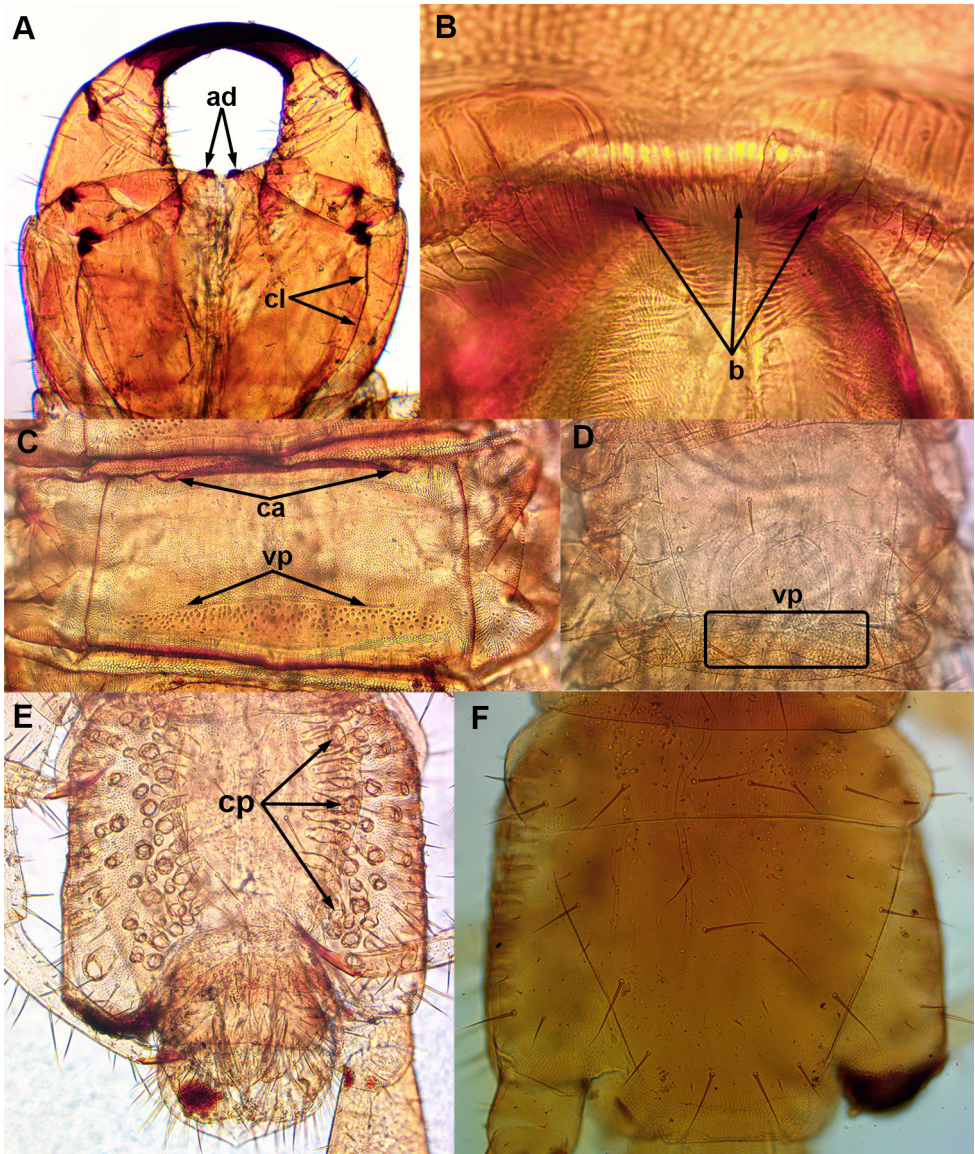
Beta diversity of studied myriapods also significantly differed between spring and summer seasons ( $p \leq 0.031$ ). Principal component analysis plots (Fig. 16) show that the observations grouped as “spring” and “summer” seasons are distant from each other, which on par with the results of PERMANOVA justifies the grouping of periods of observations and indicates that the beta diversity of studied myriapods changes with the onset of fall.

The abundance of *Brachyiulus lusitanus* ( $p=0.018$ ), *Polydesmus mediterraneus* ( $p=0.047$ ), *Julus colchicus* ( $p=0.020$ ), and *Cylindroiulus placidus* ( $p=0.047$ ) was significantly higher in the spring season, while in the summer season, the abundance of Anthroleucosomatidae Gen. sp.1 ( $p=0.047$ ) was significantly dominant (Fig. 17).

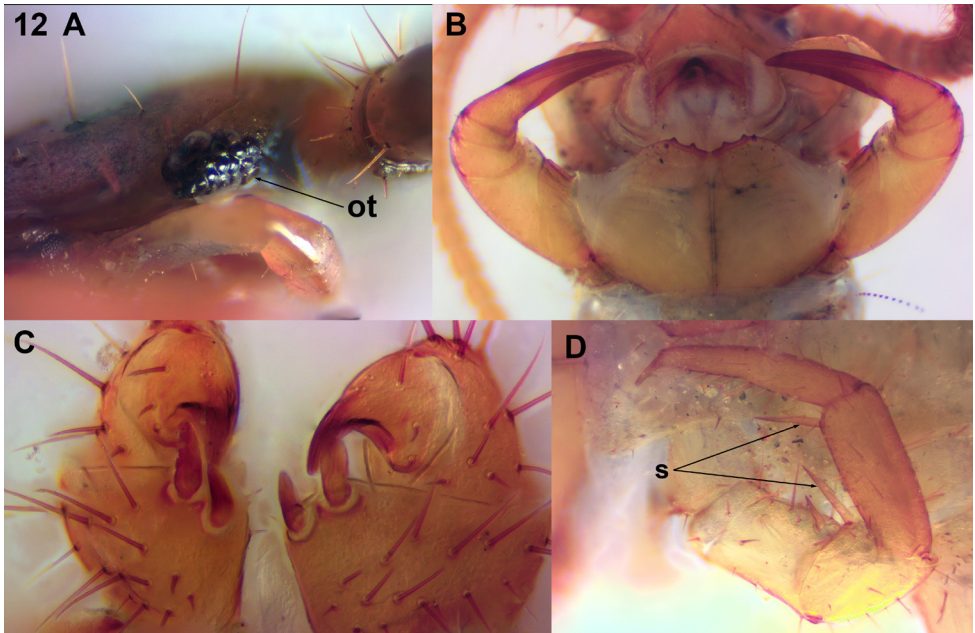
Additionally, the relative abundance of Diplopoda and Chilopoda in the studied area was investigated and their ratio was compared. As a result, there were significant differences in the Chilopoda to Diplopoda abundance ratio ( $p=0.043$ ) accompanied by an increase in the abundance of chilopods and a decrease in the abundance of diplopods in the summer season (Fig. 18).



**Figure 10.** *Harpolithobius cf. anodus* Latzel, 1880: habitus, dorsal view (A), forcipular segment, ventral view (B), leg 1 (C), tibia of leg 14 (D), tibia of leg 15 (E), dorsal views. Taken not to scale or 10 mm (A).



**Figure 11.** *Clinopodes* cf. *latisternus* (Attems, 1947): forcipular segment (A), labrum (B), metasternite of 15th leg-bearing segment (C), metasternite of 67th leg-bearing segment (D) ventral views, ultimate leg-bearing segment, ventral (E) and dorsal (F) views. Taken not to scale. Abbreviations: ad – anterior denticles, b – bristles, ca – carpophagus pit, cl – chitin lines, cp – coxal pores, vp – ventral pore field.



**Figure 12.** *Harpolithobius* sp.: head, lateral view (A), forcipular segment, ventral view (B), female gonopods, ventral view (C), leg 1 (D). Taken not to scale. Abbreviations: ot – organ of Tömösvary, s – spurs.

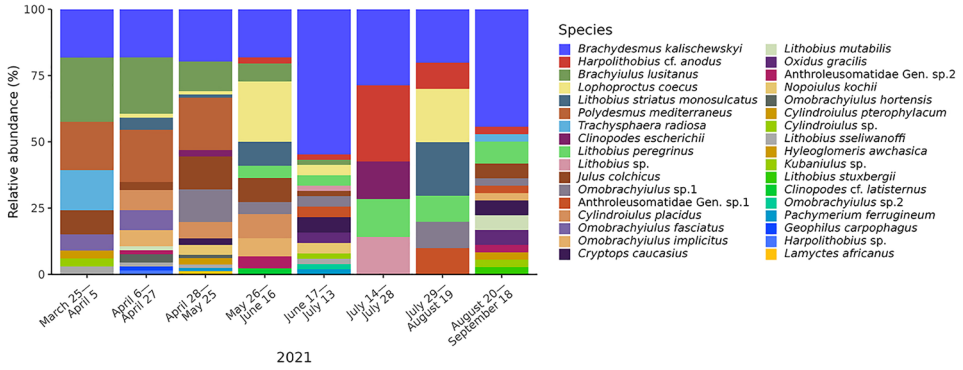


**Figure 13.** *Lithobius* sp.: habitus, dorsal view (A), tibia of 15 leg, inner view (B). Scale bars: 10 mm (A), 0.1 mm (B).

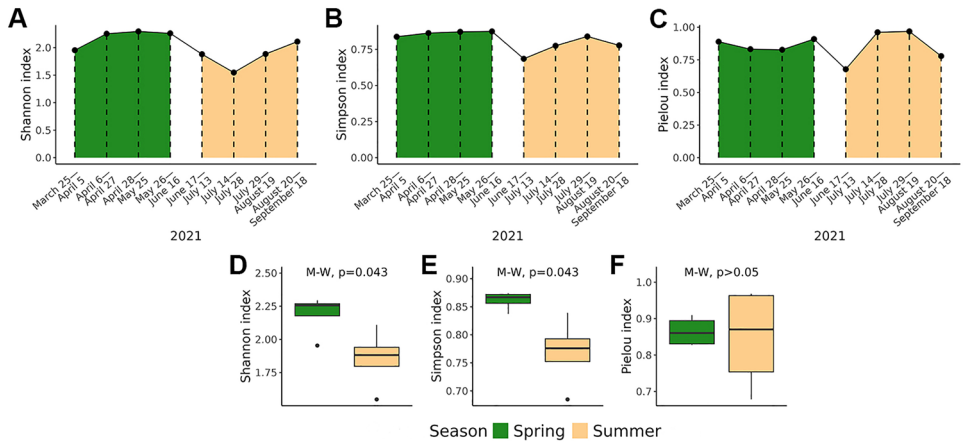
## Discussion

The diversity of myriapods in the Southern Cultures Park was low compared to natural ecosystems. For example, in a yew-boxwood grove in the same area, the class Diplopoda alone is represented by 26 species (Evsyukov et al. 2025).

Using pitfall traps, 34 species of myriapods were collected, including 20 Diplopoda and 14 Chilopoda species. Hand collecting allowed for the collection of an additional 2 millipede and 7 centipede species.



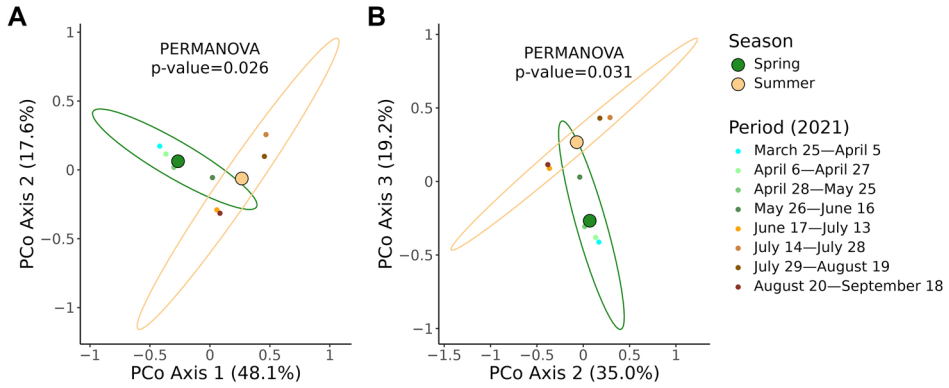
**Figure 14.** The relative abundance of recorded Myriapoda species in the Southern Cultures Park (Krasnodar Province, Southern Russia) according to the data obtained using pitfall traps between March 25, 2021, and September 18, 2021.



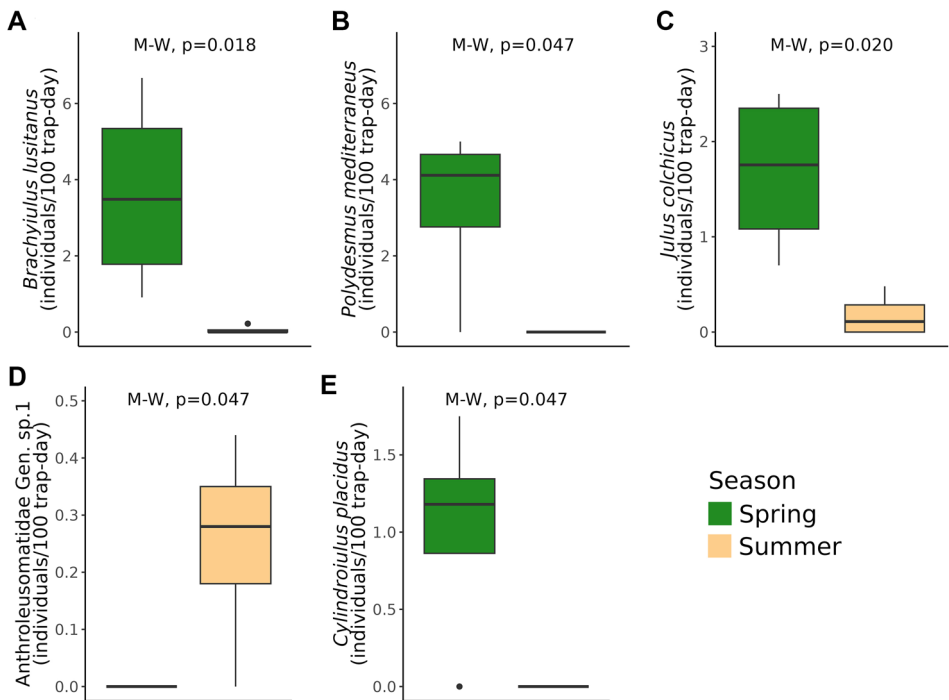
**Figure 15.** Alpha diversity of studied myriapods in the Southern Cultures Park (Krasnodar Province, Southern Russia) according to the data obtained using pitfall traps between March 25, 2021, and September 18, 2021: Shannon index (A and D), Simpson index (B and E), Pielou index (C and F). M-W=Mann-Whitney test.

Most of the diplopod species recorded are endemic or subendemic to the Caucasus, and only 6 of them have a wide distribution (Table 1). Of these, 4 species (*Brachyiulus lusitanus*, *Cylindroiulus truncorum*, *Oxidus gracilis*, and *Polydesmus mediterraneus*) can be considered invasive, as they are mainly distributed in anthropogenic habitats and were probably introduced.

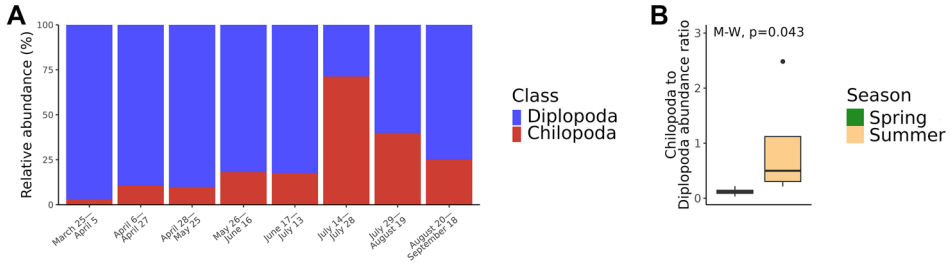
The species we recorded as *Kubaniulus* sp., found in fall and represented by a single female, is probably *K. gracilis* Lohmander, 1936, included in the Red Data Book of the Republic of Adygea (Evsyukov 2022). The unidentified species of the family Anthroleucosomatidae may also be local endemics (Antić and Makarov 2016).



**Figure 16.** Beta diversity of studied myriapods in the Southern Cultures Park (Krasnodar Province, Southern Russia) according to the data obtained using pitfall traps between March 25, 2021, and September 18, 2021: Bray–Curtis index (A), Jaccard index (B). PCoA=principal component analysis.



**Figure 17.** Myriapoda species, whose relative abundance significantly differs between spring and summer seasons: *Brachyiulus lusitanus* (A), *Polydesmus mediterraneus* (B), *Julus colchicus* (C), *Cylindroiulus placidus* (D), and Anthroleucosomatidae Gen. sp.1 (E). M-W=Mann–Whitney test.



**Figure 18.** Relative abundance of Diplopoda and Chilopoda (A), and Chilopoda to Diplopoda abundance ratio (B) in the Southern Cultures Park (Krasnodar Province, Southern Russia) according to the data obtained using pitfall traps between March 25, 2021, and September 18, 2021. M-W= Mann–Whitney test.

The picture is different for the chilopods. Of the 21 species found, only 4 are endemic or subendemic to the Caucasus (Table 1). Two species, *Harpolithobius* sp. and *Lithobius* sp., should probably be added to this list.

The remaining 15 species have wider ranges, and at least 7 of them (*Clinopodes flavidus*, *Clinopodes* cf. *latisternus*, *Geophilus carpophagus*, *Lamyctes africanus*, *L. coeculus*, *Harpolithobius* cf. *anodus*, *Lithobius forficatus*) can be classified as anthro-pochores.

**Table 1.** Chorotypes and sampling techniques of myriapods in the Southern Cultures Park

Species/subspecies	Chorotype	Sampling techniques	
		Pitfall traps	Hand sampling
Class Diplopoda			
Order Polyxenida			
Family Lophoproctidae			
<i>Lophoproctus coecus</i> Pocock, 1894	M	+	+
Order Glomerida			
Family Glomeridae			
<i>Hyleoglomeris awchasica</i> (Brandt, 1840)	CE	+	-
<i>Trachysphaera radiosa</i> (Lignau, 1911)	CE	+	-
Order Julida			
Family Blaniulidae			
<i>Nopoiulus kochii</i> (Gervais, 1847)	A	+	+
Family Julidae			
<i>Brachyiulus lusitanus</i> Verhoeff, 1898	A	+	+
<i>Cylindroiulus placidus</i> (Lignau, 1903)	CE	+	+
<i>Cylindroiulus pterophylacum</i> Read, 1992	CE	+	-

Species/subspecies	Chorotype	Sampling techniques	
		Pitfall traps	Hand sampling
<i>Cylindroiulus truncorum</i> (Silvestri, 1896)	A	-	+
<i>Julus colchicus</i> Lohmander, 1936	CE	+	-
<i>Kubaniulus</i> sp.	CE	+	-
<i>Omobrachiulus fasciatus</i> Vagalinski, 2021	CE	+	-
<i>Omobrachiulus hortensis</i> (Golovatch, 1981)	CE	+	-
<i>Omobrachiulus implicitus</i> (Lohmander, 1936)	CE	+	+
<i>Omobrachiulus</i> sp. 1	?	+	-
<i>Omobrachiulus</i> sp. 2	?	+	-
Order Polydesmida			
Family Polydesmidae			
<i>Brachydesmus furcatus</i> Lohmander, 1936	CE	-	+
<i>Brachydesmus kalischewskyi</i> Lignau, 1915	CSe	+	+
<i>Polydesmus mediterraneus</i> Daday, 1889	M	+	-
Family Paradoxosomatidae			
<i>Oxidus gracilis</i> (C.L. Koch, 1847)	A	+	+
Order Chordeumatida			
Family Anthroleucosomatidae			
Gen. sp. 1	?	+	-
Gen. sp. 2	?	+	-
Class Chilopoda			
Order Geophilomorpha			
Family Geophilidae			
<i>Clinopodes escherichii</i> (Verhoeff, 1896)	M	+	+
<i>Clinopodes flavidus</i> C.L. Koch, 1847	EM	-	+
<i>Clinopodes</i> cf. <i>latisternus</i> (Attems, 1947)	M?	+	-
<i>Diphyonyx conjungens</i> (Verhoeff, 1898)	M	-	+
<i>Geophilus carpophagus</i> Leach, 1816	EM	+	-
<i>Geophilus</i> cf. <i>flavus</i> De Geer, 1778	EM	-	+
<i>Pachymerium ferrugineum</i> (C.L. Koch, 1835)	TRa	+	+
<i>Henia illyrica</i> Meinert, 1870	M	-	+
Order Scolopendromorpha			
Family Cryptopidae			
<i>Cryptops caucasius</i> Verhoeff, 1934	CSe	+	+
Order Lithobiomorpha			
Family Henicopidae			
<i>Lamyctes africanus</i> (Porat, 1871)	A	+	-
<i>Lamyctes coeculus</i> (Brölemann, 1889)	A	-	+



Species/subspecies	Chorotype	Sampling techniques	
		Pitfall traps	Hand sampling
Family Lithobiidae			
<i>Harpolithobius</i> cf. <i>anodus</i> Latzel, 1880	EM	+	-
<i>Harpolithobius</i> sp.	?	+	-
<i>Lithobius forficatus</i> (Linnaeus, 1758)	A	-	+
<i>Lithobius mutabilis</i> L. Koch, 1862	EM	+	+
<i>Lithobius peregrinus</i> Latzel, 1880	M	+	-
<i>Lithobius sectilis</i> (Zalesskaja, 1976)	CE	-	+
<i>Lithobius striatus monosulcatus</i> Folkmanová, 1958	CE	+	-
<i>Lithobius stuxbergii</i> Sseliwanoff, 1881	CSe	+	+
<i>Lithobius sseliwanoffi</i> Garbowski, 1897	M?	+	+
<i>Lithobius</i> sp.	?	+	-

Chorotypes, from wider to increasingly narrower distributions: A — cosmopolitan/subcosmopolitan anthropochore; TPa — trans-Palaeartic; EM — Euro-Mediterranean; M — Mediterranean; CSe — sub-endemic to the Caucasus; CE — endemic to the Caucasus.

In 2021, the endemic species *Brachydesmus kalischewskyi* was among the dominant species during the quantitative observations. Only in the spring season, the second dominant diplopod species was the Eastern Mediterranean *Polydesmus mediterraneus* and was not encountered in the summer season. Among chilopods, *Harpolithobius* cf. *anodus* had the highest abundance in the summer season (Fig. 14, Suppl. material 1: Table). In general, when comparing the abundance of the two classes of myriapods by season, diplopods are represented by a greater number of species and abundance during the spring season, while chilopods are more abundant and diverse during the summer season.

In conclusion, research should be continued for possible identification of species and their population dynamics in order to determine the role of the Southern Cultures Park in the conservation of rare and endemic Caucasian species.

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## References

- Antić DŽ, Makarov SE (2016) The Caucasus as a major hotspot of biodiversity: Evidence from the millipede family Anthroleucosomatidae (Diplopoda, Chordeumatida). *Zootaxa* 4211 (1): 1–205. <https://doi.org/10.11646/zootaxa.4211.1.1>
- Arteaga A, Malumbres-Olarte J, Gabriel R, Ros-Prieto A, Casimiro P, Sanchez AF, Albergaría IS, Borges PAV (2020) Arthropod diversity in two historic gardens in the Azores, Portugal. *Biodiversity Data Journal* 8: e54749. <https://doi.org/10.3897/BDJ.8.e54749>
- Blackmore S (2017) The future role of botanical gardens. *Tropical Plant Collections. Scientia Danica. Series B, Biologica* 6: 285–297.
- Bonato L, Iorio É, Minelli A (2011) The centipede genus *Clinopodes* C. L. Koch, 1847 (Chilopoda, Geophilomorpha, Geophilidae): reassessment of species diversity and distribution, with a new species from the Maritime Alps (France). *Zoosystema* 33 (2): 175–205. <https://doi.org/10.5252/z2011n2a3>
- Dixon P (2003) VEGAN, a package of R functions for community ecology. *Journal of vegetation science* 14: 927–930.
- Dyachkov YV, Zuev RV, Gichikhanova UA (2022) Centipedes (Chilopoda) from the Dagestan, northern Caucasus, Russia. *Ecologica Montenegrina* 52: 68–89. <https://doi.org/10.37828/em.2022.52.10>
- Evsyukov A, Golovatch S, Reip HS (2016) The millipede genus *Strongylosoma* in the Caucasus (Diplopoda: Polydesmida, Paradoxosomatidae). *Acta Societatis Zoologicae Bohemicae* 80: 7–17.
- Evsyukov AP (2022) *Kubaniulus gracilis* Lohmander, 1936. In: Zamotajlov AS (Ed.) Red Data Book of Republic of Adygeya. Part 2. Animals. Maykop, 83–84. [In Russian]
- Evsyukov AP, Chumachenko YA, Popov IV (2025) Assessing the impact of a boxwood moth (*Cydalima perspectalis*) invasion on the fauna of millipedes (Diplopoda) of the Caucasian Biosphere Nature Reserve. *Zoologicheskii Zhurnal* 104 (1): 40–51. <https://doi.org/10.31857/S0044513425010038> [In Russian]
- Evsyukov AP, Golovatch SI (2013) Millipedes (Diplopoda) from the Rostov-on-Don Region, southern Russia. *Arthropoda Selecta* 22 (3): 207–215.
- Gilgado JD, Cabanillas D, Bobbitt I (2022) Millipedes and centipedes (Myriapoda: Diplopoda, Chilopoda) in Swiss heated greenhouses, with seven species new for Switzerland. *Revue suisse de Zoologie* 129 (1): 85–101. <https://doi.org/10.35929/RSZ.0063>
- Golovatch S, Evsyukov A, Reip H (2016) The millipede family Polydesmidae in the Caucasus (Diplopoda: Polydesmida). *Zootaxa* 4085 (1): 1–51. <https://doi.org/10.11646/zootaxa.4085.1.1>
- Golovatch SI, Enghoff H (1990) The millipede *Nopoulus kochii* (Gervais, 1847) in the Caucasus. In: Striganova BR (Ed.) Fauna nazemnykh bespozvonochnykh Kavkaza. Nauka Publ., Moscow, 114–118. [In Russian]
- Jaccard P (1908) Nouvelles recherches sur la distribution florale. *Bulletin de la Société vaudoise des Sciences Naturelles* 44: 223–270.
- Kime RD, Enghoff H (2011) Atlas of European Millipedes (Class Diplopoda). Vol. 1. Orders Polyxenida, Glomerida, Platydesmida, Siphonocryptida, Polyzoniida, Callipodida,

- Polydesmida. Leiden: Co-published by Pensoft Publishers, Sofia-Moscow & European Invertebrate Survey, 282 pp.
- Kime RD, Enghoff H (2017) Atlas of European millipedes 2: Order Julida (Class Diplopoda). *European Journal of Taxonomy* 346: 1–299. <https://doi.org/10.5852/ejt.2017.346>
- Korobushkin D, Semenyuk I, Tuf I (2016) An annotated checklist of the Chilopoda and Diplopoda (Myriapoda) of the Abrau Peninsula, northwestern Caucasus, Russia. *Biodiversity Data Journal* 4: e7308. <https://doi.org/10.3897/BDJ.4.e7308>
- Lignau NG (1903) Die Myriopoden der Pontus-Küsten von Caucasus. *Zapiski Novorossiyskago obshchestva estestvoispytateley* 25: 82–149.
- Maynard L, Cadena B, Thompson T, Pence V, Philpott M, O’Neil M, Pritchard M, Glenn J, Reilly B, Hubrich J, Jenike D (2023) Local plant and insect conservation evaluated with organizational identity theory. *Journal of Zoological and Botanical Gardens* 4: 214–230. <https://doi.org/10.3390/jzbg4010019>
- Minelli A (Ed.) (2011) *Treatise on Zoology. The Myriapoda. Vol. 1.* Brill, Leiden, 538 pp. <https://doi.org/10.1163/9789004188266>
- Minelli A (Ed.) (2014) *Treatise on Zoology. The Myriapoda. Vol. 2.* Brill, Leiden, 482 pp. <https://doi.org/10.1163/9789004188273>
- Minelli A, Golovatch S (2024) Myriapods. In: Scheiner SM (Ed.) *Encyclopedia of Biodiversity*. 3rd edition. Vol. 2. Elsevier, Oxford, 490–503.
- Nefediev PS, Farzalieva GS, Efimov DA (2020) New data on Lithobiomorph centipedes (Chilopoda, Lithobiomorpha) from anthropogenic habitats of Siberia. *Far Eastern Entomologist* 418: 9–14. <https://doi.org/10.25221/fee.418.2>
- Nefediev PS, Tuf IH, Farzalieva GS (2016) Centipedes from urban areas in southwestern Siberia, Russia (Chilopoda). Part 1. Lithobiomorpha. *Arthropoda Selecta* 25 (3): 257–266. <https://doi.org/10.15298/arthsel.25.3.04>
- Nefediev PS, Tuf IH, Farzalieva GS (2017) Centipedes from urban areas in southwestern Siberia, Russia (Chilopoda). Part 2. Geophilomorpha. *Arthropoda Selecta* 26 (1): 8–14. <https://doi.org/10.15298/arthsel.26.1.02>
- Neves KG (2024) Botanic gardens in biodiversity conservation and sustainability: history, contemporary engagements, decolonization challenges, and renewed potential. *Journal of Zoological and Botanical Gardens* 5: 260–275. <https://doi.org/10.3390/jzbg5020018>
- Pielou EC (1966) The measurement of diversity in different types of biological collections. *Journal of theoretical biology* 13: 131–144.
- Ponomarev AV, Chumachenko YA, Shmatko VY (2022) The first data about spider fauna (Aranei) of Dendrological Park "Yuzhnye Culture" (Adler, Krasnodar Territory, Russia). *Field Biologist Journal* 4 (2): 137–152. <https://doi.org/10.52575/2712-9047-2022-4-2-137-152> [In Russian]
- Ren H, Blackmore S (2023) The role of National Botanical Gardens to benefit sustainable development. *Trends Plant Science* 28 (7): 731–733. <https://doi.org/10.1016/j.tplants.2023.04.009>
- Shannon CE, Weaver W (1964) *The Mathematical Theory of Communication*; University of Illinois Press, Urbana, 125 pp.

- Shiryayeva NV (2019) Rare and unique collection plants of Sochi arboretum "Yuzhnyye culture", their state and ways to preserve them. *Subtropicheskoe i dekorativnoe sadovodstvo* 70: 211–222. <https://doi.org/10.31360/2225-3068-2019-211-222> [In Russian]
- Short M (2015) New records of *Lophoproctus coecus* Pocock, 1894 (Diplopoda, Polyxenida, Lophoproctidae) extend the range of the genus *Lophoproctus*. In: Tuf IH, Tajovský K (Eds) Proceedings of the 16th International Congress of Myriapodology, Olomouc, Czech Republic. *ZooKeys* 510: 209–222. <https://doi.org/10.3897/zookeys.510.8668>
- Siewers J, Schirmel J, Buchholz S (2014) The efficiency of pitfall traps as a method of sampling epigeal arthropods in litter rich forest habitats. *European Journal of Entomology* 111 (1): 69–74. <https://doi.org/10.14411/eje.2014.008>
- Simpson EH (1949) Measurement of Diversity. *Nature* 163: 688–688. <https://doi.org/10.1038/163688a0>
- Soltani GA (2014) History of the arboretum "Yuzhnye Culture" (Southern Culture) (persons and events). *Hortus Botanicus* 9: 22–33. <https://doi.org/10.15393/j4.art.2014.2241> [In Russian]
- Sorenson T (1948) A method of establishing groups of equal amplitude in plant sociology based on similarity of species content, and its application to analysis of vegetation on Danish commons. *Kongelige Danske Videnskabernes Selskab. Biologiske Skrifter* 5: 1–5.
- Stoep P, Zapparoli M, Golovatch S, Enghoff H, Akkari N, Barber A (2010) Myriapods (Myriapoda). Chapter 7.2. *BioRisk* 4 (1): 97–130. <https://doi.org/10.3897/biorisk.v4i0.51>
- Vagalinski B, Golovatch SI (2021) The millipede tribe Brachyiulini in the Caucasus (Diplopoda, Julida, Julidae). *ZooKeys* 1058: 1–127. <https://doi.org/10.3897/zookeys.1058.68628>
- Vagalinski B, Golovatch SI (2021) The millipede tribe Brachyiulini in the Caucasus (Diplopoda, Julida, Julidae). *ZooKeys* 1058: 1–127. <https://doi.org/10.3897/zookeys.1058.68628>
- Vagalinski B, Lazányi E (2018) Revision of the millipede tribe Brachyiulini Verhoeff, 1909 (Diplopoda: Julida: Julidae), with descriptions of new taxa. *Zootaxa* 4421 (1): 1–142. <https://doi.org/10.11646/zootaxa.4421.1.1>
- Volkova JS (2016) An annotated catalogue of Geophilomorph centipedes (Myriapoda, Geophilomorpha) of European Russia. *Entomological Review* 96: 500–511. <https://doi.org/10.1134/S0013873816040138>
- Wickham H (2016) ggplot2. *Elegant graphics for data analysis*. Springer Cham, 260 pp. <https://doi.org/10.1007/978-3-319-24277-4>
- Zalesskaja NT (1978) Identification book of the lithobiomorph centipedes of the USSR (Chilopoda, Lithobiomorpha). Nauka, Moscow, 211 pp. [In Russian]
- Zuev RV (2016) Centipedes (Chilopoda) from the Stavropol Territory, northern Caucasus, Russia. *Arthropoda Selecta* 25 (1): 23–38. <https://doi.org/10.15298/arthsel.25.1.03>

## Supplementary material 1

### **Table 1S. Results of sampling using pitfall traps in the Southern Cultures Park in 2021 (individuals per 100 traps per day)**

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Data type: table

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